Sensitivity analysis of populations of cardiac models in healthy and diseased states

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The normal heart contracts and relaxes at a regular beat



The myocardium is a muscle composed of tightly packed cardiomyocytes beating as a syncytium



Cardiomyocyte electrical activity can be recorded in vitro



M. Lemoine et al., Scientific Reports, 2017



Y. Xie et al., Circulation, 2015



The heart contracts because of the propagation of electrical signals throughout the myocardium in the form of <u>action potentials</u> (AP)



Single cell models describe in mathematical terms the biological processes that occur during electrical activation of a cardiomyocyte



Computational models of the heart allow to simulate the cardiac function in normal and diseased conditions

<u>Simulate aspects of electrophysiology</u>, such as APs, intracellular ionic concentrations, dynamic behavior



- Study mechanisms of initiation and maintenance of disease
- Test new treatment strategies
- Interpret experimental findings
- Simulate experimental conditions/protocols, sometimes not feasible in the lab.
- Reduce animal experiments

In **Atrial Fibrillation** is the fast irregular beating of the atria due to chaotic electrical activity



Atrial Fibrillation is a heart condition that causes the heart to beat in a fast irregular rhythm

Most common form of irregular heartbeat, or arrhythmia

Affects 10 million people in Europe and 2.5 million people in the US

Increases the risk of heart failure, stroke and death

AF is a progressive disease



Electrical & Structural

Remodelling

Cardiac alternans are correlated to AF episodes in patients and could serve as a marker for arrhythmia

Cardiac alternans

Alternans precede AF



AP: action potential \rightarrow APD alternans CaT: calcium transient \rightarrow CaT alternans



Narayan et al., Circulation, 2011

Main goal:

To use populations of models and sensitivity analysis to find which parameters of a single cell atrial model are causing the observed APD alternans behavior

The Koivumäki model of the human atrial cell showed APD alternans under dynamic pacing



- Spatial model with detailed calcium handling system
- Human ionic currents

J. van den Brink, Master Thesis, 2016

AP duration (APD) depends on the pacing cycle length (PCL)



The relationship between APD and pacing rate is represented through restitution curves



APD restitution can be used to characterize APD alternans



APD: Action potential duration

What are the cellular and molecular mechanisms underlying APD alternans in single cell models?



Large number of model instances that represent responses under different physiological conditions Explore the role of individual model parameters in the observed model behavior

Populations of models simulate different model outputs



Incorporate natural **variability** observed in experimental data

Simultaneously varying selected model **parameters** related to electrophysiology mechanisms (eg, ion channels, transporters, ionic buffering, etc.)

Simulate electrical activity and extract output measure, such as AP duration (APD), AP amplitude, resting membrane potential, APD restitution curves, etc. Parameters in population are calibrated by restricting biomarkers to within certain acceptable ranges



Data driven (eg, standard deviation)

Experimental values

Restrain the parameter space



Muszkiewicz, A et al, 2014

Sensitivity analysis is useful for understanding the behaviour of a cell model



Multivariate Linear Regression method



Multivariate Linear Regression method

$$Y_k = \beta_{0k} + \beta_{1k} \cdot x_{m1} + \beta_{2k} \cdot x_{m2} + \dots + \beta_{pk} \cdot x_{mp}$$

$$\mathbf{Y} = \mathbf{X} \cdot \mathbf{B} + \boldsymbol{\epsilon} \implies \begin{array}{l} \mathbf{X} \to \text{Model parameters} \\ \mathbf{Y} = \mathbf{X} \cdot \mathbf{B}, \\ \mathbf{Y} \to \text{Biomarkers} \end{array} \begin{array}{l} \mathbf{B} = (\mathbf{X}'\mathbf{X})^{-1}\mathbf{X}'\mathbf{Y} \end{array}$$

 $\mathbf{B} \rightarrow \text{Regression coefficients (amount of variability in response explained by the parameter)}$

B represents the amount of variability in Y that is explained by each of the parameters x when considered to be **independent.**

Alternans markers based on dynamic APD restitution



Alternans markers based on dynamic APD restitution





Baseline of Normal and AF models



Populations of Normal and AF models



Populations of Normal and AF models



Functional Calibration

Based on experimental data on human atrial cell by Sánchez et al (2014)

> APD90 APA RMP Upstroke velocity

Maximum and minimum APD in restitution curves

	SR		cAF	
	Minimum Value	Maximum Value	Minimum Value	Maximum Value
APD ₉₀ (ms)	190	440	140	330
APD ₅₀ (ms)	6	200	30	180
APD ₂₀ (ms)	1	60	1	75
APA (mV)	75	120	80	130
RMP (mV)	-85	-65	-85	-65
V ₂₀ (mV)	-35	10	-30	20
dV/dt _{max} (V/s)	40	420	40	420

Sánchez et al (2014)

Sensitivities of AP and CaT markers of normal population were consistent with literature



APD restitution of Normal cells revealed greater propensity to APD alternans compared to the AF cells



Alternans markers in Normal population



Alternans markers in Normal population showed known parameter dependencies



Alternans markers in Normal population provided new insights into role of IKur, IK1 and INaK



AF population showed highest sensitivities of alternans markers to IK1, IKur, ICaL and INaK



Normal and AF populations showed different sensitivities of alternans markers



Alternans pro-arrhythmicity

Positive correlation Negative correlation

Results

ICaL, INa, IK1, IKur AP duration INa, INaK, IK1 AP amplitude Resting membrane potential IK1 INa, IK1, INaK Upstroke velocity Calcium transient (CaT) amplitude ICaL, Ito, RyRtauinact NCX, INaK Diastolic calcium level ICaL, IKur, INaL, INaK, CaT time to peak **RyRtauact** INCX, INaK CaT time constant

Alternans pro-arrhythmicity

	Alternans threshold	ICaL, IK1 and IKur (AF), INaK, RyRmax
		and RyRtauinact (nSR)
Results	Alternans range	ICaL, INaK, RyRtauinact (nSR)
	Alternans area	ICaL (nSR), IK1, INaK, RyRtauinact
		(nSR)
Positive correlation Negative correlation	ΔAPD _{max}	ICaL, IK1, IKur, RyRtauinact (nSR)

Publication in Chaos

A computational framework for testing arrhythmia marker sensitivities to model parameters in functionally calibrated populations of atrial cells

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Conclusions and insights

Populations of Normal cells showed higher variability and propensity for APD alternans than the AF population

Normal and AF populations showed differences in the sensitivities of alternans to gCaL, gK1, gKur, and INaK conductance

Framework developed is a useful tool for studying mechanisms of cardiac alternans in single cells, and can be extended to tissue/organ simulations

This methodology can be applied to study other electrophysiology mechanisms related to arrhythmia

Future work

Incorporate variable interactions in the regression model, and reduce error by defining different a priori distributions of the parameters.

Perform simulations on populations of models in 2D tissues to uncover relationships between alternans at the cellular and tissue level

Simulate drug binding on the populations of models to simulate the effect drug therapies

Derive an arrhythmia score based on cellular biomarkers that works as a surrogate of pro-arrhythmic risk.

Thank you





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Jussi Koivumäki